


PHYS871 Clinical Imaging Applications / MIASMA

PHYS871 Clinical Imaging Applications

MIASMA

Dr Steve Barrett  April 2016

Introduction

Microscopy Image Analysis Software for Medical Applications

What is MIASMA?
A brief description of some of the projects
A more detailed look at two of the projects

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MIASMA

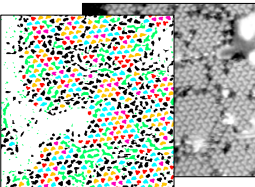
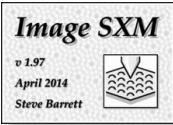
So what can a physicist do to make an impact in medicine?

Background in nanoscale physics

Expertise in image analysis of scanning microscopy images (STM, AFM, SEM)

Recognising molecular shapes (adsorption geometry)

Identifying molecular positions (substrate registration)



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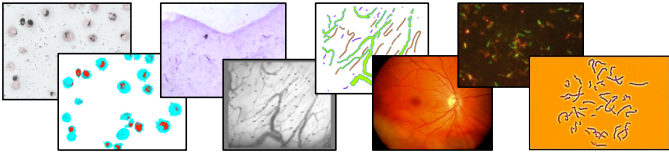

MIASMA

Liverpool Medical Imaging Network (LMI-Net) workshops

Put me in touch with medics who had image analysis problems

Some researchers within UoL, some clinicians in hospitals

Resulted in a number of collaborations



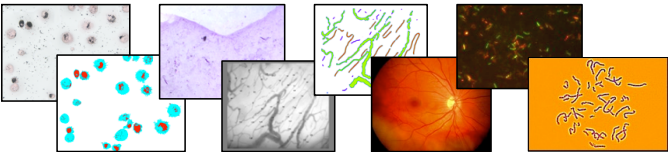
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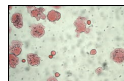
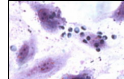
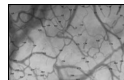

MIASMA

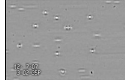
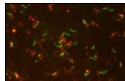
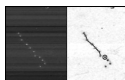
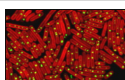
Projects include...

- Carbon particulate matter in lung cells (lung cancer)
- Parasite analysis (malaria)
- Blood flow velocities in capillary networks (meningitis)
- Retinal image analysis (diabetes)
- Parasite morphology and development (leishmania)
- Assessing antibiotic treatments (tuberculosis)



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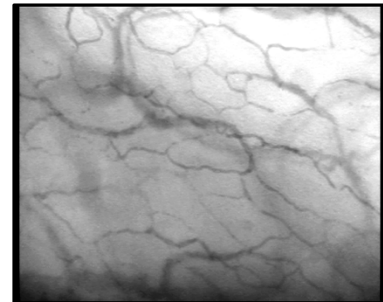
<p>Intracellular Air Pollution Particulates</p> 	<p>Collaborators Dr Stephen Gordon Liverpool School of Tropical Medicine Dr Duncan Fullerton Liverpool School of Tropical Medicine</p>	<p>Aims i) To identify particulate matter and differentiate it from cell cytoplasm. ii) To measure the area of particulate matter relative to that of the cell cytoplasm.</p> <p>Documentation MIASMA_IPA_v7.pdf</p>
<p>Malaria Parasites</p> 	<p>Collaborator Professor Alister Craig Liverpool School of Tropical Medicine</p>	<p>Aim To identify malaria parasites and differentiate them from background features.</p> <p>Documentation MIASMA_PCv5.pdf</p>
<p>Microcirculation Flow</p> 	<p>Collaborators Dr Ehtan Carol Institute of Child Health, UoL Dr Richard Sarginson Alder Hey Children's Hospital Dr Fawzia Patze UoL and Liverpool Women's Hospital</p>	<p>Aims i) To identify capillaries in videos of capillary networks and measure capillary vessel density. ii) To measure blood flow speed as a function of capillary diameter.</p> <p>Documentation MIASMA_MCA_v5.pdf</p>
<p>Retinal Imaging</p> 	<p>Collaborators Professor Simon Harding Ophthalmology Research Unit, UoL Dr Yalin Zheng Ophthalmology Research Unit, UoL</p>	<p>Aims To identify specific features such as: Blood vessel network Optic disc Haemorrhages Exudates</p> <p>Documentation Not yet available</p>

<p>Lymphocyte Flow</p> 	<p>Collaborator Dr Carlo Laudanna Department of Pathology University of Verona</p>	<p>Aims i) To identify lymphocyte cells flowing through a glass capillary. ii) To measure the length of time that cells are arrested by or rolling along the capillary wall.</p> <p>Documentation MIASMA_LF_v4.pdf</p>
<p>Bacilli Lipid Bodies</p> 	<p>Collaborator Dr Derek Sloan Clinical Sciences, UoL</p>	<p>Aim To measure the number of bacilli that contain lipid bodies.</p> <p>Documentation Not yet available</p>
<p>Fibrillin Microfibrils</p> 	<p>Collaborator Dr Riaz Avitzar Optical Biomechanics Group School of Engineering, UoL</p>	<p>Aim To speed up the analysis of microfibrils by semi-automating the process of identifying microfibril beads and calculating their xy coordinates.</p> <p>Documentation MIASMA_MFB_v3.pdf</p>
<p>Bacterial MicroCompartments</p> 	<p>Collaborator Dr Luning Liu Institute of Integrative Biology, UoL</p>	<p>Aim To determine the locations of microcompartments within the outlines of bacterial membranes.</p> <p>Documentation Not yet available</p>

Case Study 1 : Microcirculation Analysis

Take one MIASMA project as an example...

Blood flow velocities in capillary networks (meningitis)

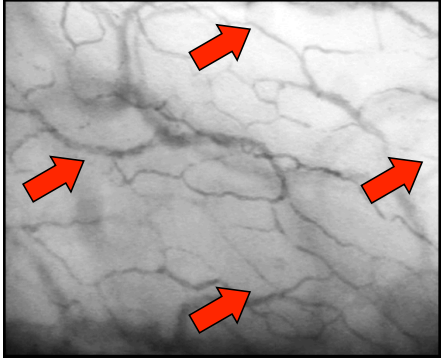


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Video Stability

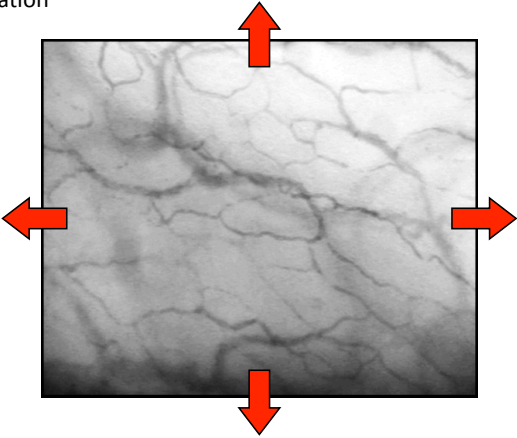
Translation



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Video Stability

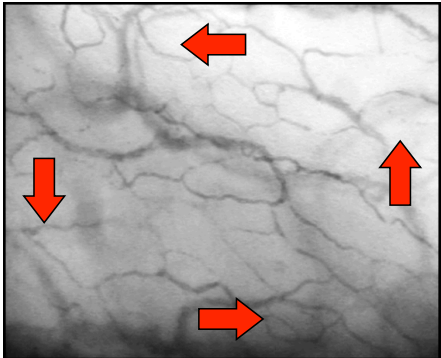
Magnification



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Video Stability

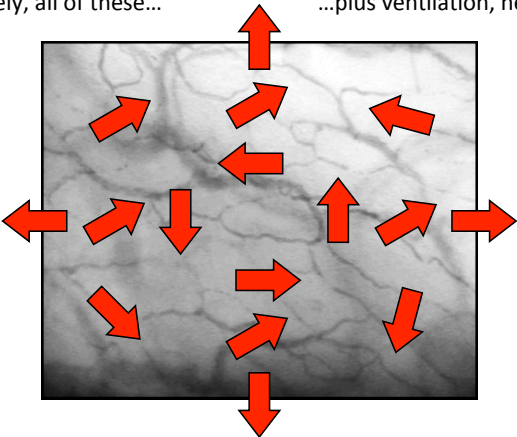
Rotation



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Video Stability

More likely, all of these... ...plus ventilation, heartbeat



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Microcirculation Analysis

What information can be extracted?

How should the microcirculation be quantified?

What (manual) scoring systems exist?

Percentage of perfused vessels (PPV)
(Perfused = flow exists for > 50% of the time)

Microcirculation Flow Index (MFI)
(Is the flow 'intermittent' or 'sluggish' or OK?)

Microcirculation Analysis

Calculation of blood flow speeds

- **Stabilisation** of the video
- Identification of the blood **vessels** (which are invisible)
- **Isolation** of each capillary vessel
- Analysis of the **movement** of the blood cells

Quantification of the flow distribution (PPV and MFI)

- Flow speed as a function of **time**
- Flow speed as a function of vessel **diameter**
- **Variations** in flow speeds across the vessel network

Microcirculation Analysis

Calculation of blood flow speeds

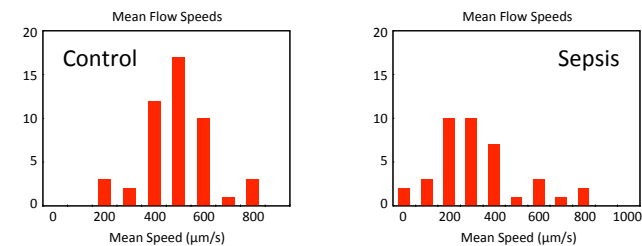
- **Stabilisation** of the video << **Fourier Methods**
- Identification of the blood **vessels** << **Kernel Filters**
- **Isolation** of each capillary vessel << **Particle Analysis**
- Analysis of the **movement** of the blood << **Fourier Methods**

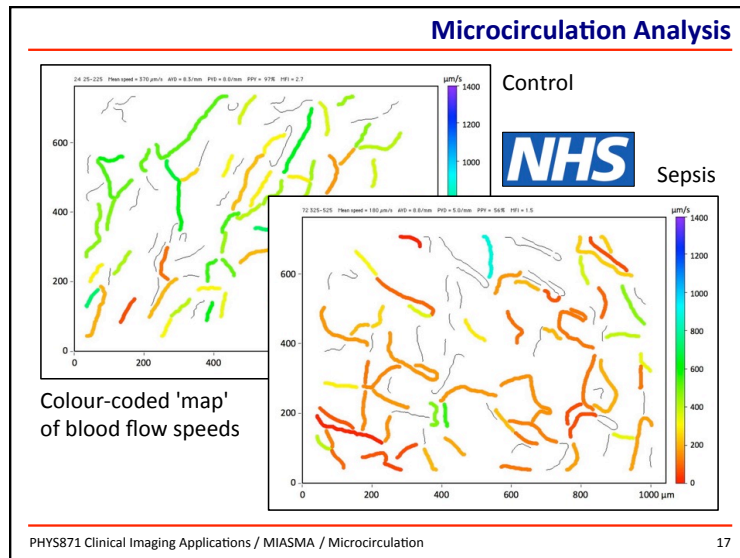
Quantification of the flow distribution (PPV and MFI)

- Flow speed as a function of **time**
- Flow speed as a function of vessel **diameter**
- **Variations** in flow speeds across the vessel network

Microcirculation Analysis

Through a combination of techniques, including cross-correlations (to stabilise the video images) and autocorrelations (to identify the motion of blood cells that are barely detectable) it is possible to quantify the blood flow speeds in vessels as small as 7 μm diameter.





Case Study 2 : Investigating Cancer

This final section will cover the preliminary results of the research carried out under the EPSRC critical mass grant

"Disease diagnosis through spectrochemical imaging of tissues"
(Weightman, Martin, Barrett + Cockcroft, Lancaster, Manchester, Cardiff)

Roughly speaking, that translates to...

Can we identify an infrared absorption signature for tissue that is likely to become cancerous?

Or...

Can we detect cancer before it is cancer?

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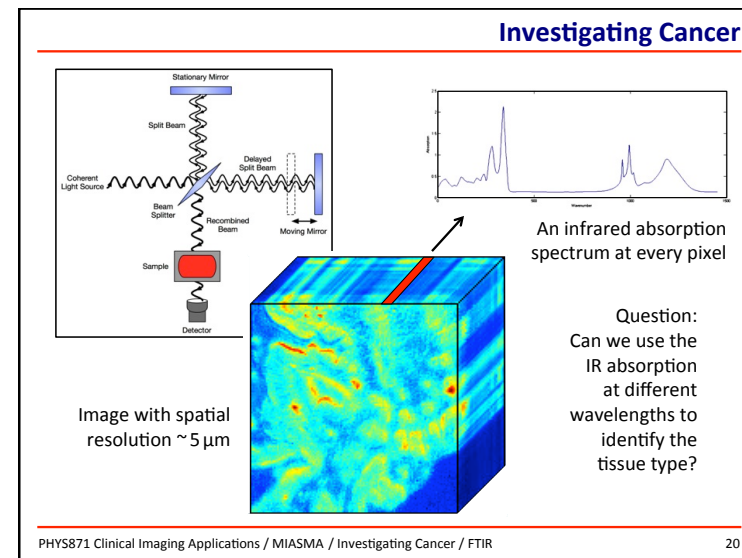
Investigating Cancer

What tissues are being studied?

We started with oesophageal cancer, and its precursor called Barrett's oesophagus (no relation, as far as I am aware):

A condition in which the tissue lining the oesophagus is replaced by tissue that is similar to the intestinal lining (intestinal metaplasia). People with Barrett's oesophagus have an increased risk for developing oesophageal cancer.

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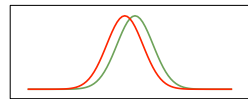


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Investigating Cancer

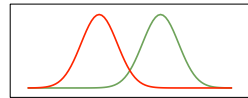
In general, infrared absorption at different wavelengths is very similar even for different tissue types. So, what wavelengths should we use to discriminate one (abnormal and potentially cancerous) tissue type from another (normal and healthy) type?

Certain pairs of wavelengths are much better than others, and they're not necessarily the ones we would have guessed by looking at the spectra.



Poor choice of λ 's

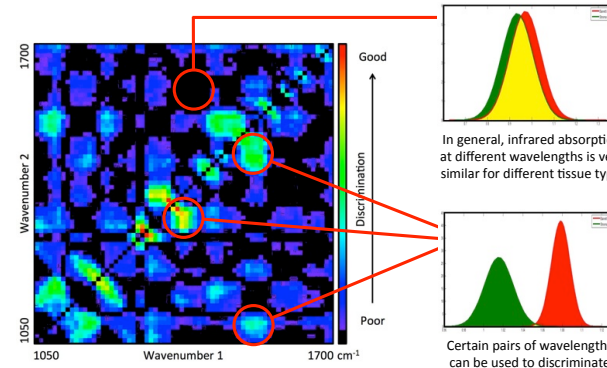
Making histograms of the ratios of the values of IR absorption at different wavelengths shows this very clearly.



Good choice of λ 's

Histograms of ratios of IR absorption for abnormal (red) and normal (green) tissue

Investigating Cancer



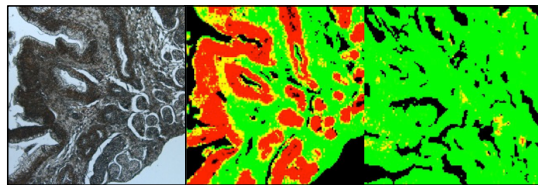
"Butterfly diagram"

In general, infrared absorption at different wavelengths is very similar for different tissue types

Certain pairs of wavelengths can be used to discriminate abnormal from normal tissue

Investigating Cancer

Selecting the best discrimination from the butterfly diagram, we can generate a map identifying different tissue types.



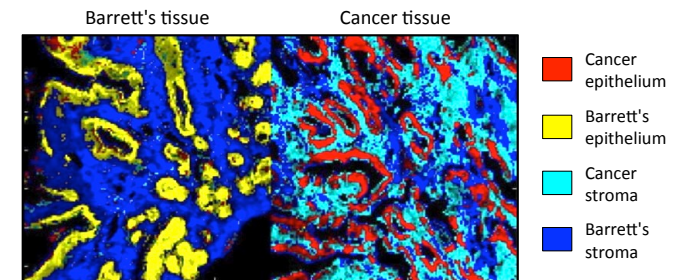
Visible image

Abnormal (red) and normal (green)

Normal tissue only

Investigating Cancer

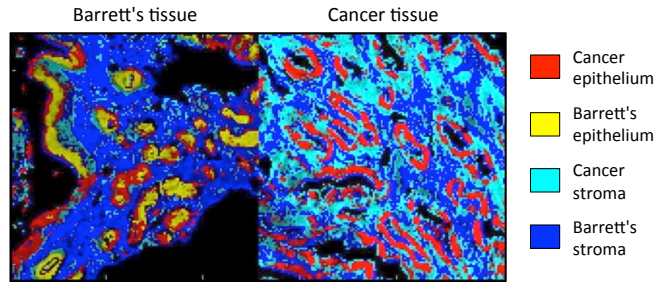
This idea was then extended to identify more than two tissue types...



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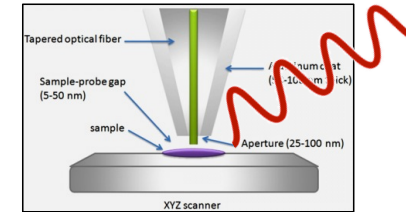
Investigating Cancer

... and then tested on tissues not used to 'train' the analysis routine.



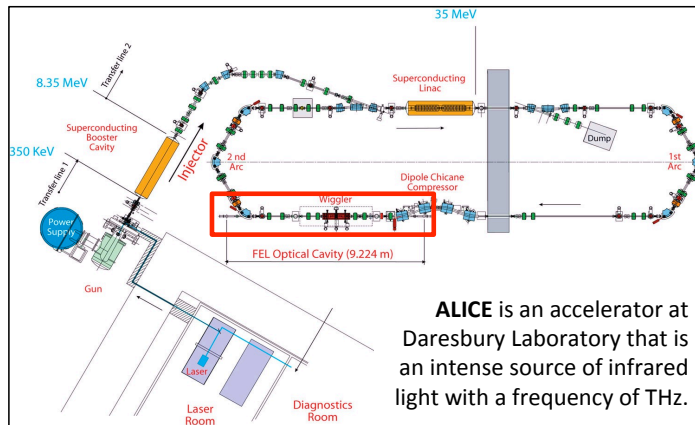
Investigating Cancer

To improve the spatial resolution we need to beat the diffraction limit using Scanning Near-Field Optical Microscopy (SNOM).



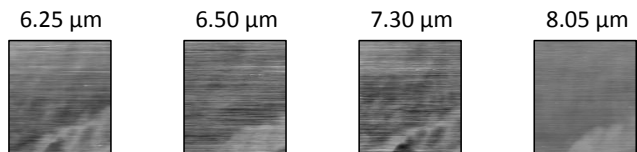
Imaging with sub- μm resolution requires plenty of infrared photons to illuminate the sample underneath the scanning tip. This is where a free-electron laser that operates in the infrared comes in.

Free-Electron Laser

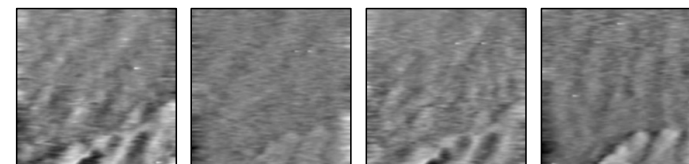


ALICE is an accelerator at Daresbury Laboratory that is an intense source of infrared light with a frequency of THz.

SNOM Imaging



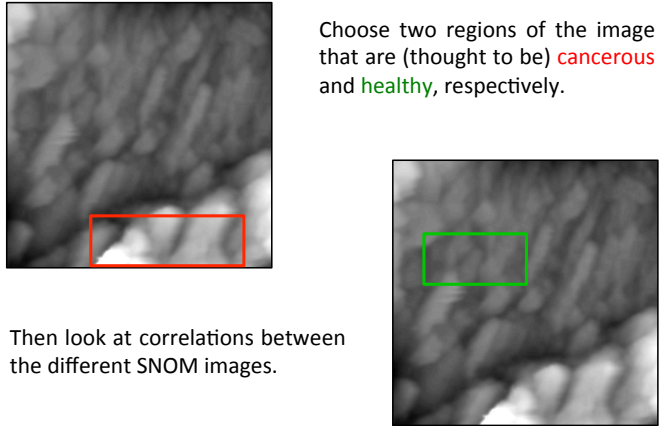
Raw images as acquired by the SNOM at different IR wavelengths



Processed to remove artefacts and make features easier to see

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Image Correlation

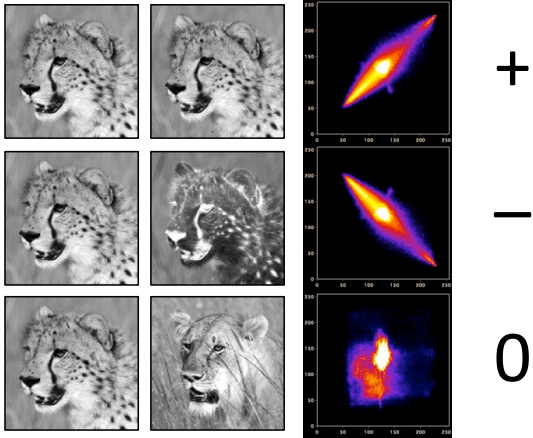


Choose two regions of the image that are (thought to be) **cancerous** and **healthy**, respectively.

Then look at correlations between the different SNOM images.

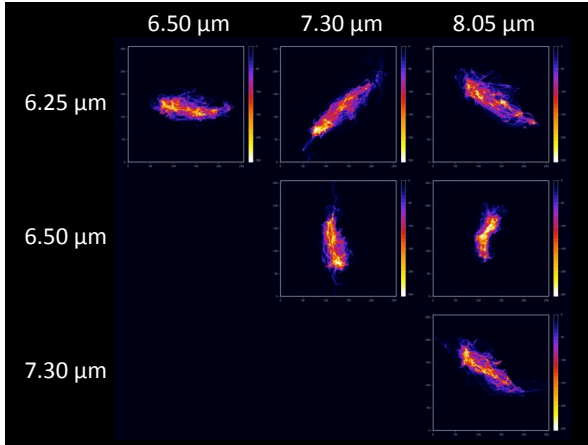
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Image Correlation



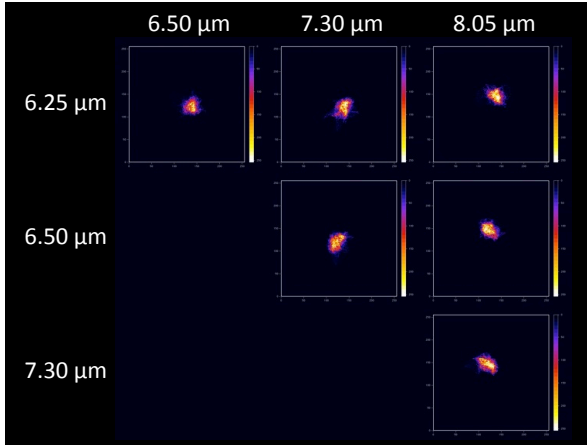
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Image Correlation – Cancer



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Image Correlation – Healthy



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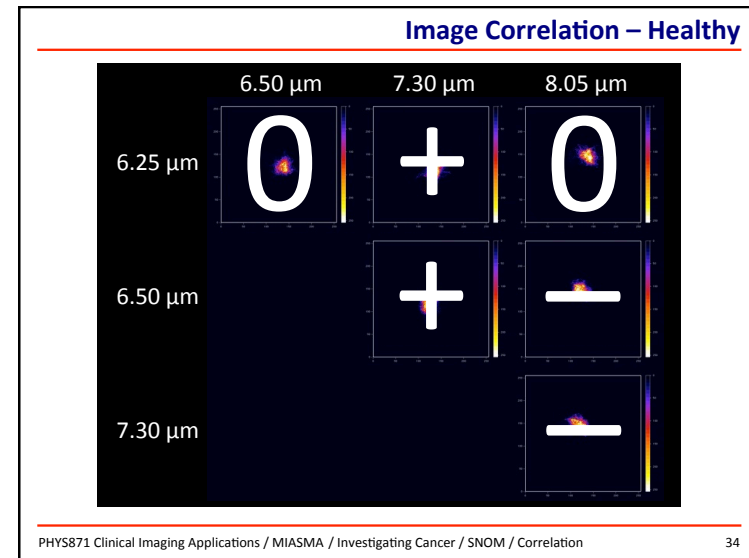
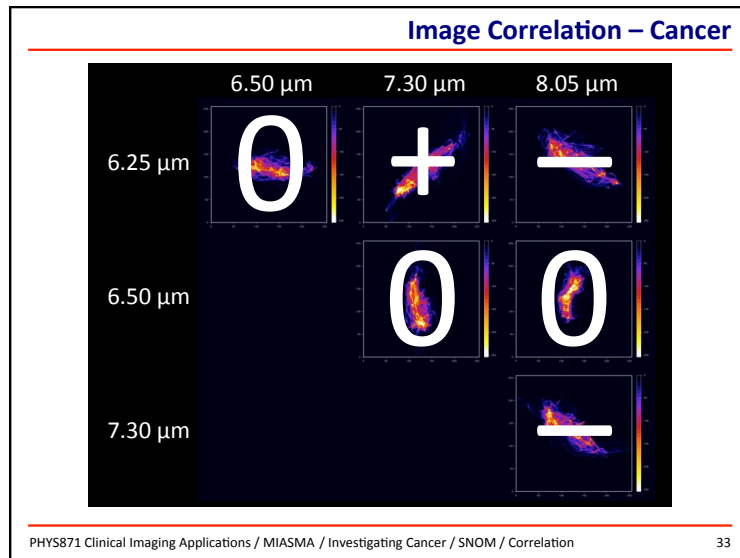


Image Correlation

Can the patterns of correlations between images taken at different wavelengths provide the 'signatures' of cancerous, pre-cancerous and healthy tissue?

0	+	-
0	0	-

⇒ Cancer ?

0	+	0
+	-	-

⇒ Healthy ?

The research is still in the early stages, but the results of the analysis to date indicates that we have found a technique and a method of analysis that has the potential to do just that.

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Acknowledgements

Thanks to

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